

NoToxRox
10 Midelton Ave
North Bondi
NSW 2026



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Accreditation Number 1261
Site Number 1254

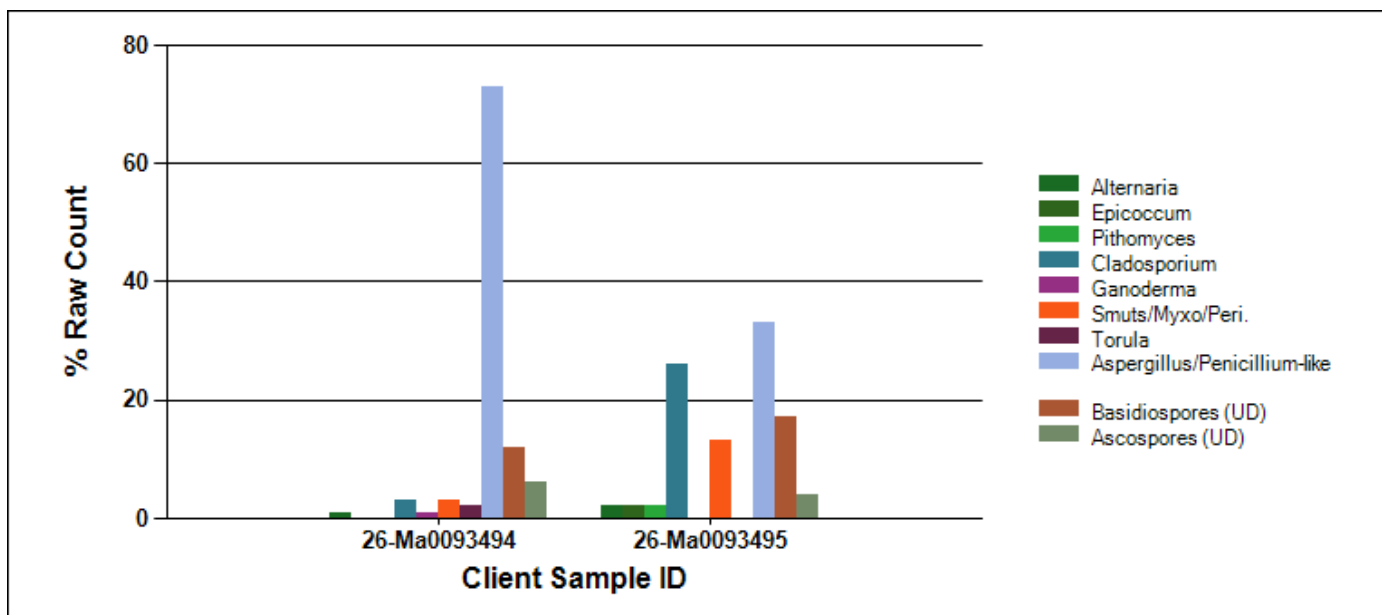
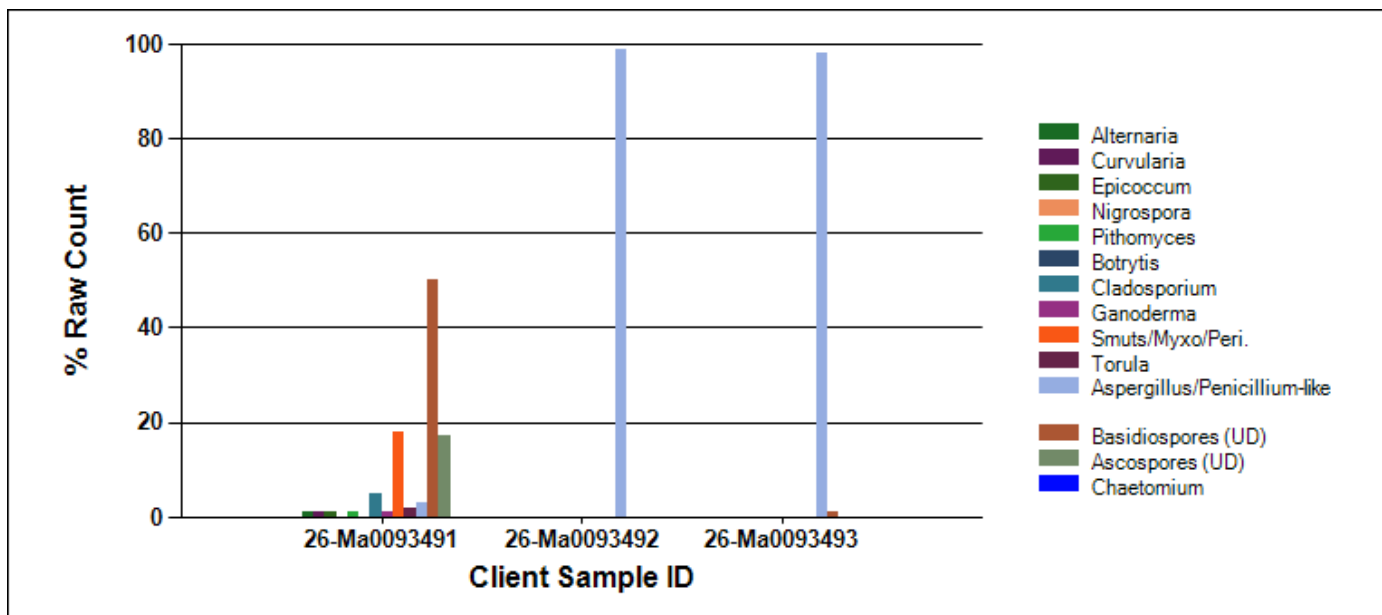
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inspection, proficiency testing scheme providers and
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Attention: **Joanne Lia**

Report **1341343-ML**
Project name **9 WILDTHORN AVE DURAL NSW**
Project ID **DURAL**
Received Date **Mar 31, 2026**

Client Sample ID	OUTSIDE			LOWER MESSAGE ROOM			OFFICE			ENTRANCE		
Sample Matrix	Spore Trap			Spore Trap			Spore Trap			Spore Trap		
Eurofins Sample No.	26-Ma0093491			26-Ma0093492			26-Ma0093493			26-Ma0093494		
Date Sampled	Mar 27, 2026			Mar 27, 2026			Mar 27, 2026			Mar 27, 2026		
Spore Trap Mould Analysis												
% Analysed	100			100			100			100		
Flow Rate (L/min)	15			15			15			15		
Sampling time (min)	5			5			5			5		
LOR	13			13			13			13		
Background Debris	2			3			3			3		
	Raw Counts	fs/m ³	%	Raw Counts	fs/m ³	%	Raw Counts	fs/m ³	%	Raw Counts	fs/m ³	%
Hyphal Fragments	8	110		6	80		5	67		2	27	
Un-ID	-	-	-	-	-	-	-	-	-	-	-	-
Identification												
Pollen	2	27		4	53		1	13		-	-	
<i>Alternaria</i>	1	13	1	2	27	< 1	-	-	-	1	13	1
<i>Arthrinium</i>	-	-	-	-	-	-	-	-	-	-	-	-
Ascospores (UD)	18	240	17	5	67	< 1	13	170	< 1	7	93	6
<i>Aspergillus/Penicillium</i> -like	3	40	3	M21 2100	28000	99	M21 2800	37000	98	93	1200	73
Basidiospores (UD)	52	690	50	9	120	< 1	15	200	1	15	200	12
<i>Drechslera/Bipolaris</i> -like	-	-	-	-	-	-	-	-	-	-	-	-
<i>Botrytis</i>	-	-	-	-	-	-	1	13	< 1	-	-	-
<i>Chaetomium</i>	-	-	-	1	13	< 1	-	-	-	-	-	-
<i>Cladosporium</i>	5	67	5	2	27	< 1	4	53	< 1	4	53	3
<i>Curvularia</i>	1	13	1	1	13	< 1	-	-	-	-	-	-
<i>Epicoccum</i>	1	13	1	2	27	< 1	-	-	-	-	-	-
<i>Ganoderma</i>	1	13	1	-	-	-	-	-	-	1	13	1
<i>Nigrospora</i>	-	-	-	-	-	-	1	13	< 1	-	-	-
<i>Pithomyces</i>	1	13	1	3	40	< 1	1	13	< 1	-	-	-
Smuts/Myxomycetes/Periconia	19	250	18	5	67	< 1	10	130	< 1	4	53	3
<i>Stachybotrys/Memnoniella</i>	-	-	-	-	-	-	-	-	-	-	-	-
<i>Torula</i>	2	27	2	-	-	-	-	-	-	2	27	2
Total (fs/m³)	104	1400		2130	28000		2845	38000		127	1700	

Client Sample ID	LIVING - AERIAL		
Sample Matrix	Spore Trap		
Eurofins Sample No.	26-Ma0093495		
Date Sampled	Mar 27, 2026		
Spore Trap Mould Analysis			
% Analysed	100		
Flow Rate (L/min)	15		
Sampling time (min)	5		
LOR	13		
Background Debris	3		
	Raw Counts	fs/m³	%
Hyphal Fragments	3	40	
Un-ID	-	-	-
Identification			
<i>Alternaria</i>	1	13	2
<i>Arthrinium</i>	-	-	-
Ascospores (UD)	2	27	4
<i>Aspergillus/Penicillium</i> -like	15	200	33
Basidiospores (UD)	8	110	17
<i>Drechslera/Bipolaris</i> -like	-	-	-
<i>Chaetomium</i>	-	-	-
<i>Cladosporium</i>	12	160	26
<i>Curvularia</i>	-	-	-
<i>Epicoccum</i>	1	13	2
<i>Nigrospora</i>	-	-	-
<i>Pithomyces</i>	1	13	2
Smuts/Myxomycetes/Periconia	6	80	13
<i>Stachybotrys/Memnoniella</i>	-	-	-
Total (fs/m³)	46	620	



Sample History

Where samples are submitted/analysed over several days, the last date of extraction is reported.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

Description

Spore Trap Mould Analysis

- Method: AEML-DOC-18: Spore Trap Analysis

Testing Site

Melbourne

Extracted

Mar 31, 2026

Holding Time

N/A

Internal Quality Control Review and Glossary

General

- QC data may be available on request.
- Samples were analysed on an 'as received' basis.
- Information identified in this report with blue indicates data provided by the customer, which may impact the results.
- This report replaces any interim results previously issued.
- Spores of *Aspergillus*, *Penicillium*, and others are small with few distinguishing features and, therefore, can be challenging to differentiate.
- If % analysed is < 100%, spores per m³ is based on extrapolation and not actual count.
- # Estimation performed due to high count.

Holding Times

Please refer to the 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least six hours prior to sample receipt deadlines as stated on the Sample Receipt Advice (SRA).

If the Laboratory did not receive the information in the required timeframe, and despite any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the sampling date; therefore, compliance with these may be outside the laboratory's control.

Units

Total Fungal Count	Raw count of fungal structures in slide area analysed	g	gram
fs/m³	fungal structures per cubic metre	%	Percentage
fs/cm²	fungal structures per square centimetre		

Terms

<	Less than	-like	Spores lacking distinguishable characteristics from other similar spores
>	Greater than	LOR	Limit of Reporting
% Analysed	The amount of the trace that was analysed	N/A	Not Applicable
COC	Chain of Custody	UD	Undifferentiated
fs	fungal structures - a collective term for a fragment or groups of fragments from fungi, including but not limited to conidia, conidiophores, hyphae and spores	PLM	Polarising Light Microscope
Hyphal Fragments	Hyphae, mycelia or fruiting bodies – fragmented or intact	Raw Counts	The number of spores counted by the analyst
Intact Hyphae	Hyphal structures where conidiophores are still attached to conidia.	Un-ID	Unidentified fungal particulate

Water Indicator

Fungi are most commonly associated with indoor mould growth in buildings with long-term water intrusion issues.

Background Debris: Background debris is the amount of non-fungal particulate present in the trace, including dust, fibres, skin cells, dust mites, and insect parts. A debris rating is assigned to each trace from 0 (lowest) to 5 (highest). A higher debris rating means samples are more challenging to analyse, and spores, especially smaller spores like *Aspergillus/Penicillium*, may be obscured. The degree of bias increases with the percentage field of view occluded. A further explanation of the debris rating is listed below:

- No particulate matter was observed.
- Minimal. > not observed to approximately 5% particulate matter observed. Reported values are minimally affected by particle load.
- Approximately 5% to 25% of the field of view occluded with particulate matter. Negative bias is expected.
- Approximately 25% to 75% of the field of view is occluded with particulate matter. Negative bias is expected.
- Approximately 75% to 90% of the field of view is occluded with particulate matter. Negative bias is expected.
- Greater than 90% of the field of view is occluded with particulate matter. Accurate quantification is not possible due to a large negative bias

When an estimate is performed, the estimated count lies between the following ranges

>100	Count lies between 100 & 199	>2500	Count lies between 2500 & 4999
>200	Count lies between 200 & 499	>5000	Count lies between 5000 & 7499
>500	Count lies between 500 & 999	>7500	Count lies between 7500 & 9999
>1000	Count lies between 1000 & 1999	>10000	Count is greater than 10000

Indoor and Outdoor Comparisons

There are no current industrial standards regarding permissible levels of airborne fungi that may be present in buildings. It is common for fungal spores to be present in a normal indoor environment. A general guideline widely accepted in the industrial hygiene industry is that the types and numbers of mould spores present in the indoor environment should be similar to those in the outdoor environment. If inside spore counts are significantly higher than outside counts, this may indicate a potential mould problem. The comparison of outdoor and indoor spore types and concentrations is a valuable tool in assessing abnormal mould contamination; however, it should not be the sole determining factor in evaluating health risks and remediation strategies.

Combustion Products

This is a semi-qualitative analytical method. The relative percentage of combustion-by-products such as char, ash, and soot is visually estimated and reported as a percentage using direct microscopy technique. Other opaque particulates, such as paint, rubber, metal oxides, etc., are also reported as a combined percentage describing the major constituents.

^^The soot identification by PLM is presumptive.

Analytical results are not corrected for field and analytical blanks. Test results relate only to the items tested and cannot be extrapolated to anything more significant than their original intent. Interpretation of the analytical results is the sole responsibility of the customer.

Comments**Sample Integrity**

Custody Seals Intact (if used)	N/A
Sample correctly preserved	N/A
Appropriate sample containers have been used	Yes
Some samples have been subcontracted	No

Qualifier Codes/Comments

Code	Description
M21	Count is an estimate only

Authorised by:

Catherine Wilson Analytical Services Manager
Kalysha Murphy Senior Analyst-Mould



Glenn Jackson
Managing Director

Final Report – this report replaces any previously issued Report

- Indicates Not Requested

* Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request

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